

EXHIBIT 6

Expression Characteristics of Two Potential T Cell Mediator Genes

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Received January 17, 1989; accepted March 29, 1989

Lymphocyte subset-specific cDNA clones were recently isolated by a modified differential screening procedure. The expression patterns of two of these cDNAs, designated as 4-1BB and L2G25B, were studied in greater detail. Nucleotide sequence comparison revealed that 4-1BB was not previously recognized. Although the L2G25B sequence had been recognized recently, the function of the encoded molecule has yet to be well studied. The transcripts of the two cDNAs were inducible by concanavalin A in mouse spleen cells, cloned helper T cells (L2), cloned cytotoxic T cells (I.3), and cytolytic T cell hybridomas. They were also inducible with stimulation through antigen receptor (TCR), with immobilized anti-TCR antibody in cloned T cells L2, dB45, and L3. Concanavalin A inducibility was inhibited by cyclosporin A. They were not inducible by IL-2 stimulation. The expression patterns of these transcripts were similar to those of IFN- γ , except that the level of transcripts of the two cDNAs was at least fivefold lower than that of IFN- γ , and the peak level of expression occurred earlier. These data suggest that L2G25B and 4-1BB may represent new T cell mediators. © 1999 Academic Press, Inc.

INTRODUCTION

T lymphocytes play a central role in the immune network both as effectors and regulators. They are composed of subsets endowed with distinct helper, suppressor, and cytolytic capabilities. These functions may be mediated by surface receptors and subset-specific immune effectors which are elaborated and secreted after stimulation either with lectin or specific antigen or immobilized monoclonal antibody (mAb) against T cell antigen receptor (TCR) (1-4). The genes for a number of the subset-specific T cell effector molecules have been cloned, but not all activities are correlated with the cloned genes. Identification and demonstration of such unrecognized molecules can uncover hitherto-unknown functions of T cells.

This laboratory has recently cloned a series of T cell subset-specific cDNAs from cloned helper T (HTL) L2 and cloned cytolytic T (CTL) L3 cells by employing a modified differential screening procedure (5). Nucleotide sequences of two cDNA clones, L2G25B and 4-1BB, were determined (6). The deduced amino acid sequences revealed that both contain putative leader sequences. The protein encoded by 4-1BB had a potential membrane anchor segment and other features also seen in known receptor proteins (6). A human homolog of L2G25B was reported (7) and the same

mouse sequence has been isolated. The characteristics of expression of L2 and L3 are the focus of the study of inducibility and expression.

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Cells. Cloned murine CTL^a 1⁺, L3T4⁻, and H-2L^d reactive 1⁺, Lyt-2⁻, L3T4⁺, and Mls^{a/c} PN37 are derived by the fusion (11). They are Thy-1⁺, LFA-1⁺, IL-2R⁺, and CD45R⁺.

Md90, PN37, BW5147, and lin A (5 μ g/ml) at a cell concentration of 10⁶/ml. Md90 and PN37 cells was monitored for production. Mouse thymocyte growth factor/nophorhol-13-acetate (TPA), 20 hr. Stimulation was monitored in C57BL/6, BALB/c, or Swiss Webster mice. Navalalin A (5 μ g/ml) at a cell concentration of 10⁶/ml. K46 (17), rat NK cell LGL (18), and B cells were not stimulated with any of the above.

Isolation of T cell-specific cDNAs. cDNAs that are specific for T cells were isolated by negative differential screening as described previously (1). T cell-specific cDNAs were further analyzed by sequencing. Two cDNAs, L2G12 and L2G25B, were reported elsewhere (1). A third cDNA, L2G25, was isolated recently from a lymphocyte cell line (1). The sequence was isolated recently from a macrophage cell line RAW264.7 (8).

RNA blot hybridization. Total RNA was fractionated on 1.2% agarose-formaldehyde gels (BRL, Bethesda, MD; or New England Nuclear, Boston, MA), transferred to nylon membranes, and used for probe labeling and hybridization as described previously (20). Probes were labeled by random priming with 32 P-dCTP (Ampliscript, BRL) and used at 1–2 ng/ml. The filters were washed at room temperature for 1 h in 0.1× SSC and 0.1% SDS, and then for 1 h in 0.1× SSC and 0.1% SDS at 65°C.

mouse sequence has been isolated from the murine macrophage cell line (8). The characteristics of expression of the two cDNA clones which were expressed in both L2 and L3 are the focus of the present report. These clones, because of their patterns of inducibility and expression, may represent potential new T cell mediators.

MATERIALS AND METHODS

Cells. Cloned murine CTL L3 (9) and dB45 cells (10) are Thy-1,2⁺, Lyt-2⁺, LFA-1⁺, L3T4⁺, and H-2L^d reactive. Cloned murine HTL L2 cells (9) are Thy-1,2⁺, LFA-1⁺, Lyt-2⁺, L3T4⁺, and Mls^{a/d} reactive. The cytolytic T cell hybridomas Md90 and PN37 are derived by the fusion of BW5147 thymoma and BALB/c antiEL-4 CTL (11). They are Thy-1⁺, LFA-1⁺, Lyt-2⁺, L3T4⁺, and H-2D^b reactive.

L2 cells were stimulated with concanavalin A (10 µg/ml) for 14 hr, or with immobilized anti-TCR mAb F23.1 (12) for 6 hr at a cell concentration of 10⁶-10⁷/ml. L3 cells were stimulated with concanavalin A (2 µg/ml) for 14 hr, or with immobilized clonotypic anti-TCR mAb 384.5 (13) for various time periods, or with recombinant human IL-2 (100 µg/ml) (Cetus Corp., Emeryville, CA) for 6 hr at a cell concentration of 2.5 × 10⁶/ml. In other experiments, L3 cells were treated with concanavalin A alone, concanavalin A plus cyclosporin A (0.2 µg/ml), or concanavalin A plus actinomycin D (1 µg/ml) for 6 hr. L2, L3, and dB45 cell preparations were kindly provided by David Lancki at the University of Chicago, Illinois.

Md90, PN37, BW5147, and CTLLA11 (14) cells were stimulated with concanavalin A (5 µg/ml) at a cell concentration of 5 × 10⁶ cells/ml for 4 hr. Stimulation of Md90 and PN37 cells was monitored by increased cytotoxicity and increased IL-2 production. Mouse thymoma EL-4 cells (15) were stimulated with 12-O-tetradecanoylphorbol-13-acetate (TPA, 10 ng/ml) at a cell concentration of 1.0 × 10⁶/ml for 20 hr. Stimulation was monitored by IL-2 assay (16). Splenocytes were obtained from C57BL/6, BALB/c, or Swiss Webster mouse spleens and were stimulated with concanavalin A (5 µg/ml) at a cell concentration of 5.0 × 10⁶/ml for 14 hr. B cell lymphoma K46 (17), rat NK cell LGL (18), and mouse melanoma, Cloudman S-91 (19) cells were not stimulated with any of the above reagents.

Isolation of T cell-specific cDNA clones. We have previously isolated a group of cDNAs that are specific for T cells in contrast to B cells, employing both positive and negative differential screening and RNA blot analysis of various lymphoid cells. The T cell-specific cDNAs were further studied to determine whether they were specific for cloned HTL L2 or cloned CTL L3. The identity of these cDNA sequences, and nucleotide and deduced amino acid sequences of previously unrecognized 4-1BB and L2G25B, were reported elsewhere (6). We learned, however, that the L2G25B sequence was isolated recently from lipopolysaccharide (LPS)-stimulated murine macrophage cell line RAW264.7 (8).

RNA blot hybridization. Total cytoplasmic RNA or poly(A)⁺ RNA was fractionated on 1.2% agarose-formaldehyde gels and transferred to Gene Screen Plus (New England Nuclear, Boston, MA). Gel-purified cDNA inserts were ³²P-labeled by nick translation and used as probes. Filters were prehybridized and hybridized at 42°C in 50% formamide, 5× SSC (1× SSC=150 mM NaCl, 15 mM sodium citrate, pH 7.0), 0.1% SDS, 250 µg/ml of salmon sperm DNA, and 10% dextran sulfate. Filters were washed at room temperature for 15 min in 2× SSC and 0.1% SDS, and at 42°C for 5 min in 0.1× SSC and 0.1% SDS several times. When a Northern blot of Gene Screen

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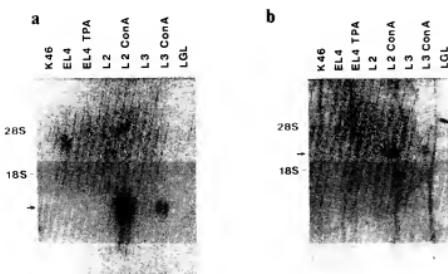


Fig. 1. T cell-specific expression of L2G25B and 4-1BB mRNA. Poly(A)⁺ mRNA was prepared from mouse B cell line (K46), unstimulated EL-4 (EL-4), TPA-stimulated EL-4 (EL-4 TPA), and rat NK cell line (LGL); and total RNA was prepared from unstimulated L2 (L2), concanavalin A-stimulated L2 (L2 Con A), unstimulated L3 (L3), and concanavalin A-stimulated L3 (L3 Con A). Ten micrograms of total RNA or ten micrograms of poly(A)⁺ RNA was fractionated on a formaldehyde/agarose gel, transferred to Gene Screen Plus, and hybridized to ³²P-labeled L2G25B (a) and 4-1BB (b) sequentially. Positions of 28S and 18S rRNA markers are indicated. An arrow indicates the specific hybridization signal.

Plus was used multiple times for hybridization, the previous probe was removed by treating the membrane in 10 mM Tris-HCl (pH 7.0), 0.2% SDS at 85° for 1 hr.

RESULTS

L2G25B and 4-1BB Are Expressed Preferentially in T Cells

L2G25B was isolated from an L2 cDNA library, and 4-1BB was isolated from an L3 cDNA library by a modified differential screening (5). As shown in Figs. 1a and 1b, L2G25B and 4-1BB were expressed preferentially in L2 and L3 cells only after concanavalin A stimulation. The sizes of transcripts were approximately 800 bases for L2G25B and 2400 bases for 4-1BB. The abundance of the two transcripts was 5- to approx 10-fold higher in L2 cells than in L3 cells. The two transcripts were not detectable in K46 B cells, EL-4 thymoma cells, or rat large granular lymphocytes. L2G25B mRNA was consistently more abundant than 4-1BB mRNA.

L2G25B and 4-1BB mRNA Are Inducible by TCR Stimulation, but Not by IL-2 Stimulation

The inducibility of the two cDNA clones was tested after L3 TCR stimulation by clonotypic anti-TCR mAb, 384.5, or IL-2. As shown in Figs. 2a and 2b, the expression of the two cDNAs was inducible by TCR stimulation, but not by IL-2 stimulation in L3 cells. L2G25B mRNA was detectable at 0.5 hr after TCR stimulation, peaked at 6 hr, and decreased thereafter until at least 24 hr. 4-1BB mRNA was detectable at a very low level in unstimulated L3 cells in this experiment. The induction of 4-1BB mRNA occurred approximately 6 hr after TCR stimulation and remained level until 24 hr.

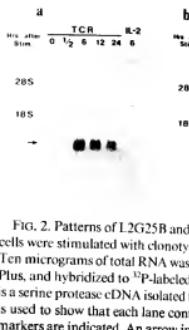


Fig. 2. Patterns of L2G25B and 4-1BB mRNA expression after TCR or IL-2 stimulation. Cells were stimulated with clonotypic TCR. Ten micrograms of total RNA was fractionated on a formaldehyde/agarose gel, transferred to Gene Screen Plus, and hybridized to ³²P-labeled cDNA. Plus, unstimulated L3 cells were stimulated with IL-2 (100 U/ml) for 0.5, 1/2, 6, 12, and 24 hr. A serine protease cDNA isolated from L3 cells was used to show that each lane contains equal amounts of total cellular RNA. Lane 6 is a serum protease cDNA isolated from L3 cells. mRNA markers are indicated. An arrow in each panel indicates the specific hybridization signal.

Figure 2c shows the kinetics of L2G25B mRNA expression after TCR stimulation used in Fig. 2a or 2b. IFN- γ peaked at 12 hr, and declined thereafter. L2G25B mRNA in unstimulated L3 cells was approximately 10-fold higher than that of 4-1BB mRNA with that of L2G25B mRNA.

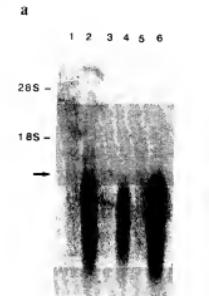


Fig. 3. Expression of L2G25B mRNA after stimulation with anti-mAb 384.5. dBA5 cells were stimulated with anti-mAb 384.5 for 6 hr. Ten micrograms of total RNA was fractionated on a formaldehyde/agarose gel, transferred to Gene Screen Plus, and hybridized to ³²P-labeled L2G25B (a). Lane 6 is a control. An arrow indicates the L2G25B mRNA band.

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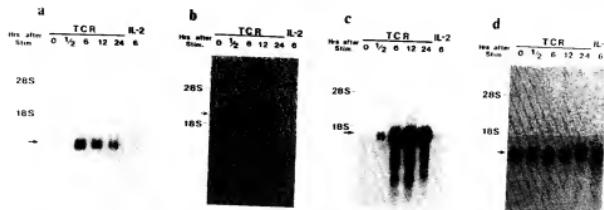


FIG. 2. Patterns of L2G25B and 4-1BB mRNA expression after TCR stimulation or IL-2 treatment. L3 cells were stimulated with clonotypic anti-TCR mAb 384.5 for 0, 1/2, 6, 12, or 24 hr, or with rIL-2 for 6 hr. Ten micrograms of total RNA was fractionated on a formaldehyde/agarose gel, transferred to Gene Screen Plus, and hybridized to ³²P-labeled L2G25B (a), 4-1BB (b), IFN- γ (c), and L3G10#6 (d) cDNA. L3G10#6 is a serine protease cDNA isolated from L3 cell cDNA library, which is identical to HF gene (29). L3G10#6 is used to show that each lane contains an almost equal amount of RNA. Positions of 28 and 18 S rRNA markers are indicated. An arrow indicates the specific hybridization signal.

Figure 2c shows the kinetics of IFN- γ mRNA expression in the same RNA blot as used in Fig. 2a or 2b. IFN- γ mRNA was detectable at 0.5 hr after TCR stimulation, peaked at 12 hr, and declined slightly until 24 hr. There was a low level of IFN- γ mRNA in unstimulated L3 cells. When we compared the peak levels of L2G25B and 4-1BB mRNA with that of IFN- γ mRNA, IFN- γ mRNA was at least 5-fold higher than that of L2G25B mRNA and at least 25-fold higher than that of 4-1BB mRNA.

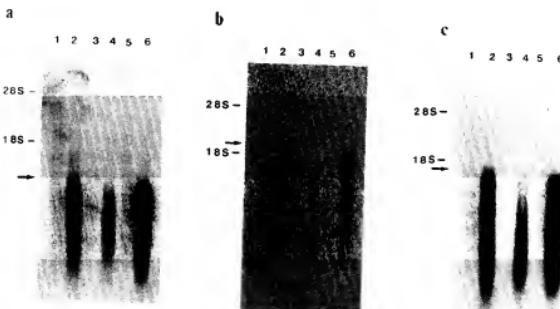


FIG. 3. Expression of L2G25B and 4-1BB mRNA in HTL L2 and CTL dB45 cells. HTL L2 and CTL dB45 cells were stimulated with anti-TCR mAb F23.1 for 6 hr. L3 cells were stimulated with anti-TCR mAb 384.5 for 6 hr. Ten micrograms of total RNA from unstimulated L3 (lane 1) and stimulated L3 (lane 2), unstimulated dB45 (lane 3), stimulated dB45 (lane 4), unstimulated L2 (lane 5), and stimulated L2 (lane 6) was fractionated on formaldehyde/agarose denaturing gel, transferred to Gene Screen Plus, and hybridized to ³²P-labeled L2G25B (a), 4-1BB (b), and IFN- γ (c) cDNA. A fraction of RNA in each lane was degraded and detected as RNA in lower molecular sizes.

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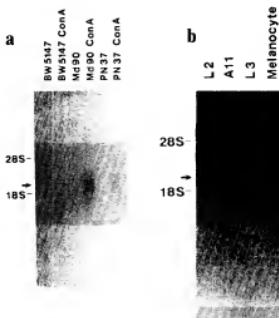


FIG. 4. Expression of 4-IBB mRNA in concanavalin A-stimulated hybridomas PN37 and Md90, and in a CTL CTLA11. (a) Ten micrograms of poly(A)⁺ mRNA from BW5147, PN37, and Md90 cells, both stimulated and unstimulated, was fractionated, transferred to nitrocellulose filter, and probed with ³²P-labeled 4-IBB cDNA probe. (b) Ten micrograms of poly(A)⁺ mRNA from mouse melanoma cells (melanocyte) and 10 µg of total RNA from unstimulated L2 (L2), L3 (L3), and stimulated CTLA11 (A11) cells were fractionated, transferred to Gcne Screen Plus, and hybridized to ³²P-labeled 4-IBB cDNA probe.

Figure 2d demonstrates that all six lanes contained almost identical amounts of RNA. The probe was a serine protease cDNA (L3G10#6) isolated from L3 cells (5). In summary, the pattern of the two cDNA expressions was similar to that of IFN- γ expression upon TCR stimulation.

L2G25B and 4-IBB mRNA Are Inducible by TCR Stimulation in Other Cloned HTL, CTL, and Hybridomas

As shown in Figs. 3a and 3b, L2G25B and 4-IBB mRNA are also inducible in HTL L2 and CTL dB45 after TCR stimulation with anti-TCR mAb F23.1. The mRNA level for the two cDNAs was also much lower than that of IFN- γ in L2 and dB45 cells (Fig. 3c). L2 cells show the highest level of expression of the three cell clones. We also found that 4-IBB mRNA was inducible by concanavalin A in two cytotoxic hybridomas, PN37 and Md90 (Fig. 4a), and a CTL line CTLA11 (Fig. 4b).

Effects of Cyclosporin A on L2G25B and 4-IBB Transcription

We next examined the effect of cyclosporin A on RNA expression of L2G25B and 4-IBB. Cyclosporin A inhibits mitogen- or antigen-induced T cell proliferation (20–22). It has also been shown to block the induction of expression of several lymphokine genes, including IL-2 and IFN- γ (23–25). The inhibition of lymphokine production occurs at a pretranslational level (24–26). In contrast, cyclosporin A appears to have no effect on the inducible expression of *c-fos* and IL-2 receptor genes in T cells (23). As shown in Figs. 5a and 5b, cyclosporin A inhibited the induced accumulation of L2G25B and 4-IBB mRNA. The same findings were seen with IFN- γ (Fig. 5c). Fig-

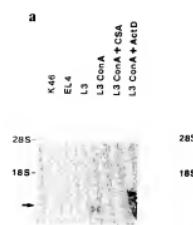


FIG. 5. Effect of cyclosporin A on L2G25B and 4-IBB mRNA expression. (a) Total RNA was prepared from untreated L3 (L3 Con A + ActD) and EL-4 cells (EL-4) and L3 cells treated with Concanavalin A plus cyclosporin A (L3 Con A + CSA). (b) L2 cells were treated with Concanavalin A plus cyclosporin A (L2 Con A + CSA). Cells were fractionated, transferred to nitrocellulose filter, and probed with ³²P-labeled L2G25B (a), 4-IBB (b), or IFN- γ (c) cDNA probe. An arrow indicates the 18S rRNA loading control.

ure 5d shows that cyclosporin A inhibits the expression of the serine protease protease (probe L3G10#6) mRNA. These data strongly suggest that L2G25B and 4-IBB activation requirements are similar.

L2G25B and 4-IBB mRNA Are Induced by TCR Stimulation in Other Cloned HTL, CTL, and Hybridomas

To find out whether or not cloned T cells or hybridomas can be stimulated with concanavalin A, we used L2 cells and L3 cells. As shown in Figs. 6a and 6b, the two mRNAs, L2G25B and BALB/c mouse thymocytes and mouse splenocytes (data not shown), were induced by concanavalin A-stimulated L2 cells. The L3G10#6 mRNA was not detectable in L3 cells (data not shown). These data suggest that L2G25B and 4-IBB mRNA are induced by appropriate stimulation.

L2G25B and 4-IBB share similar properties as follows: (1) The mRNA levels of the two genes are induced by concanavalin A or by other

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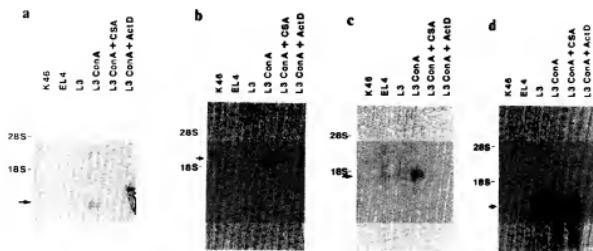


FIG. 5. Effect of cyclosporin A on L2G25B and 4-1BB mRNA expression. L3 cells were stimulated with concanavalin A, concanavalin A plus cyclosporin A, or concanavalin A plus actinomycin D. Ten micrograms of total RNA from unstimulated L3 (L3), concanavalin A-stimulated L3 (L3 Con A), concanavalin A plus cyclosporin A-treated L3 (L3 Con A + CsA), and concanavalin A plus actinomycin D-treated L3 (L3 Con A + ActD) cells and 10 μ g of poly(A)⁺ mRNA from K46 (K46) and TPA-stimulated EL4 cells (EL4) were fractionated, transferred to Gene Screen Plus membrane, and hybridized to ³²P-labeled L2G25B (a), 4-1BB (b), L3G10#6 (c), and L3IFN- γ (d) cDNA. Cyclosporin A treatment did not alter the level of L3G10#6 mRNA, but almost completely abrogated the induced expression of the other three mRNA species. An arrow indicates a specific hybridization signal.

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ure 5d shows that cyclosporin A had minimal or no effect on the level of a serine protease (probe L3G10#6) mRNA, and shows that the three lanes contained almost equal amounts of RNA (EL4 or K46 cells did not express L3G10#6 mRNA). These data strongly suggest that L2G25B and 4-1BB expression may show some of the same activation requirements as other known lymphokines.

L2G25B and 4-1BB mRNA Are Inducible in Normal Mouse Spleen Cells

To find out whether or not the expression of these genes was unique to certain cloned T cells or hybridoma cells, splenocytes from C57BL/6 and BALB/c mice were stimulated with concanavalin A and tested for mRNA expression. As shown in Figs. 6a and 6b, the two mRNAs were detectable after concanavalin A stimulation in C57BL/6 and BALB/c mouse splenocytes. They were also inducible in Swiss Webster mouse splenocytes (data not shown). As shown in Fig. 6c, IFN- γ mRNA was detectable in concanavalin A-stimulated BALB/c splenocytes (for unknown reasons, IFN- γ mRNA was not detectable in concanavalin A-stimulated C57BL/6 splenocytes in this experiment). RNA preparations for Fig. 6c were different from those for Figs. 6a and 6b. These data suggest that these molecules may be induced in normal mouse spleen cells by appropriate stimuli, as in the cloned T cells.

DISCUSSION

L2G25B and 4-1BB share properties of soluble T cell mediators. The properties are as follows: (1) The mRNAs of the two are preferentially expressed in T cells. (2) The mRNAs of the two genes are present in undetectable amounts in T cells until induced by concanavalin A or by TCR stimulation. (3) The patterns of expression are

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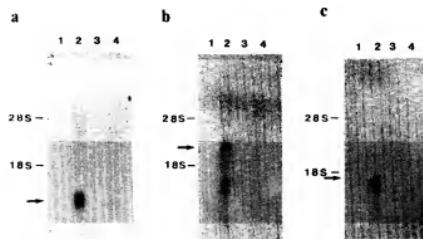


FIG. 6. Expression of L2G25B and 4-1BB mRNA in mouse splenocytes. Splenocytes were obtained from C57BL/6 and BALB/c mice and stimulated with concanavalin A for 14 hr. Ten micrograms of total RNA from unstimulated BALB/c (lane 1) and stimulated BALB/c (lane 2), unstimulated C57BL/6 (lane 3), and stimulated C57BL/6 (lane 4) splenocytes was fractionated, transferred to Gene Screen Plus, and hybridized to 32 P-labeled L2G25B (a), 4-1BB (b), and IFN- γ (c) cDNA.

very similar to that of the lymphokine IFN- γ . (4) Cyclosporin A inhibits the induced mRNA expression corresponding to the two cDNAs. In addition, sequence analysis of L2G25B showed features consistent with several analyzed lymphokine cDNAs; for example, the small size of mRNA, potential signal sequence, and repeated AUUUA element in 3' untranslated region (27). In contrast, while the 4-1BB exhibits expression patterns which resemble those of the lymphokine mRNAs, the sequence analysis of this cDNA appears consistent with those of known receptor proteins. It would be interesting, therefore, to determine the function of 4-1BB. Since Davatelas *et al.* (8) isolated the cDNA clone identical to L2G25B from the LPS-stimulated RAW264.7 cell line, we tested the mRNA expression of L2G25B and 4-1BB in the RAW264.7 cells. L2G25B mRNA was inducible within 30 min by LPS stimulation in the cells, but 4-1BB mRNA was not detectable over a 36-hr induction period. Interestingly, the induction of L2G25B mRNA was not inhibited by cyclosporin A in the RAW264.7 macrophage line (Fig. 7).

Using the same concanavalin A-stimulated L2 cells, Prystowsky *et al.* (28) identified 10 different lymphokine activities from culture supernatants. They include IL-2, IL-3, B cell stimulatory factor, granulocyte/macrophage colony-stimulating factor, IFN- γ , and five unidentified factors which affect macrophage activities. In the course of the studies, we isolated and identified cDNAs for IL-2, IL-3, GM-CSF, T cell replacing factor, and proenkephalins from our concanavalin A-stimulated L2 cDNA library (5, 6). We suggest, therefore, that L2G25B might represent the novel soluble mediators of Prystowsky *et al.* (28), which affect macrophage activities. However, the nature of the 4-1BB gene product is difficult to predict.

By applying a modified differential screening of L2 and L3 cDNA library, two novel T cell genes were isolated. The two T cell genes were expressed at low levels compared with IFN- γ expression level, and these genes shared expression properties with several of the known lymphokines. Correlation of the T cell molecules with functional activities is the next critical step.

FIG. 7. Expression of L2G25B mRNA. *Escherichia coli* 0127:B8, Siga, contains 20 μ g of total RNA from unstimulated (lane 1), stimulated with LPS + cyclosporin A (lane 2), stimulated with LPS + cyclosporin A (lane 3), and stimulated with LPS + cyclosporin A (lane 4). Electrophoresis on agarose gel, transferred to Gene Screen Plus, and hybridized to cDNA probes of 28 and 18 s mRNA markers.

The author thanks Dr. Sherman David Lancki, and Mike Prystowsky LGL; Dr. John Farrar for EL-4 and Ding-E Young for CTLLA11 cells; Cloudman S-91 cells. I also thank Dr. Asifa Haq and Thomas Savin and Mrs. Wall, and Mrs. Mary Kiefer for typing.

This work was supported in part by the Feasibility Grant Program of Am-

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DNA library, two assessed at low levels expression properties of cell molecules with

FIG. 7. Expression of L2G25B mRNA in RAW264.7. RAW264.7 cells were stimulated with 1 μ g/ml of LPS (*Escherichia coli* 0127:B8, Sigma, St. Louis, MO), or LPS + cyclosporin A (0.2 μ g/ml). Each lane contains 20 μ g of total RNA from RAW264.7 cells, unstimulated (lane 1) or stimulated with LPS for 2 hr (lane 2), stimulated with LPS + cyclosporin A for 2 hr (lane 3), stimulated with LPS for 6 hr (lane 4), or stimulated with LPS + cyclosporin A for 6 hr (lane 5). The RNA was fractionated on a formaldehyde/agarose gel, transferred to Gene Screen Plus membrane, and hybridized to 32 P-labeled L2G25B. The positions of 28 and 18 S mRNA markers are each indicated. An arrow indicates the specific hybridization signal.

ACKNOWLEDGMENTS

The author thanks Dr. Sherman Weissman for his continued advice and support; Drs. Frank Fitch, David Lancki, and Mike Prystowsky for providing the L2 and L3 cell preparations; Dr. Pierre Henkart for LGL; Dr. John Farrar for EL-4 and CT 6 cells; Dr. Charles Janeway for A20.2J and K46 cells; Dr. John Ding-E Young for CTLA11 cells; Hoffmann-LaRoche for the IFN- γ probe; and Dr. Ruth Halaban for Cloudman S-91 cells. I also thank Dr. Steve Litwin and Mrs. Mary Kiefer for editing the manuscript; Drs. Asifa Haq and Thomas Savin and Mrs. Alex Liddell for technical help; and Mrs. Helen Kelley, Mrs. Elaine Wall, and Mrs. Mary Kiefer for typing the manuscript.

This work was supported in part by Grants AI23058 and AI28175 from the National Institutes of Health, the Feasibility Grant Program of American Diabetes Association, and the Schering Corporation.

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Total Lymphoid Irradiation
and Enhances Specific

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Schoo

Revere

Thymus-independent primary
untreated and TLI-treated NZB mice
show a low primary response to *Brahma*
BALB/c mice. However, TLI
induces a strong secondary antibody response at day 14.
NZB/NZW or nonautoimmunogenic
or Ficoll were masked by high
levels in the anti-BA response, spon-
taneously decreased after TLI.

NZB/NZW F1 female mice
are susceptible to human systemic lupus erythematosus.
Antibodies, hypergammaglobulinemia,
immune complex glomerulonephritis,
and various immunologic abnormalities
are similar to human disease.

It has been reported, as the
only IgG secretion by spleen cells
of NZB mice is due to antibodies (2, 3), as well as a
vitro, in response to the poly-
purified protein derivative of
exogenous antigens is variable.

Total lymphoid irradiation
kin's disease (9), can markedly
with moderate or advanced renal
disease. One month after TLI, there is an e-

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